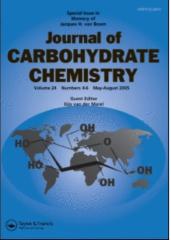
This article was downloaded by: On: 23 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



# Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713617200

Synthetic Studies on Sialoglycoconjugates 71: Synthesis of Sulfo- and Sialyl-Lewis × Epitope Analogs Containing the 1-Deoxy-*N*-acetylgulcosamine in Place of *N*-Acetylglucosamine Residue

Hirokazu Maeda<sup>a</sup>; Hideharu Ishida<sup>a</sup>; Makoto Kiso<sup>a</sup>; Akira Hasegawa<sup>a</sup> <sup>a</sup> Department of Applied Bioorganic Chemistry, Gifu University, Gifu, Japan

**To cite this Article** Maeda, Hirokazu , Ishida, Hideharu , Kiso, Makoto and Hasegawa, Akira(1995) 'Synthetic Studies on Sialoglycoconjugates 71: Synthesis of Sulfo- and Sialyl-Lewis  $\times$  Epitope Analogs Containing the 1-Deoxy-N-acetylgulcosamine in Place of N-Acetylglucosamine Residue', Journal of Carbohydrate Chemistry, 14: 3, 369 — 385

To link to this Article: DOI: 10.1080/07328309508002078 URL: http://dx.doi.org/10.1080/07328309508002078

# PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

# SYNTHETIC STUDIES ON SIALOGLYCOCONJUGATES 71: SYNTHESIS OF SULFO- AND SIALYL-LEWIS X EPITOPE ANALOGS CONTAINING THE 1-DEOXY-N-ACETYLGLUCOSAMINE IN PLACE OF N-ACETYLGLUCOSAMINE RESIDUE

Hirokazu Maeda, Hideharu Ishida, Makoto Kiso and Akira Hasegawa\*

Department of Applied Bioorganic Chemistry, Gifu University, Gifu 501-11, Japan

Received November 1, 1994 - Final Form January 4, 1995

## ABSTRACT

Sulfo and sialyl Le<sup>x</sup> epitope analogs containing the 1-deoxy-N-acetylglucosamine in place of GlcNAc residue have been synthesized. Glycosylation of 2-acetamido-1,5anhydro-3,6-di-O-benzyl-2-deoxy-D-glucitol (10) with 2,4,6-tri-O-benzoyl-3-Olevulinyl- $\alpha$ -D-galactopyranosyl trichloroacetimidate (6) prepared from 2-(trimethylsilyl)ethyl  $\beta$ -D-galactopyranoside (1) in five steps, or with the  $\alpha$ -sialyl-(2 $\rightarrow$ 3)galactose donor, afforded the desired  $\beta$ -glycosides 12 and 15, respectively. Glycosylation of the acceptor 14 derived from 12 in two steps, or 16 derived from 15 by reductive removal of the benzyl group followed by selective O-benzoylation, with 2,3,4-tri-O-benzyl-1-thio-β-L-fucopyranoside methyl (17), using dimethyl(methylthio)sulfonium triflate, gave the trisaccharide 18 and tetrasaccharide 23. Compound 18 was converted via reductive removal of the benzyl groups, Oacetylation, selective removal of the levulinyl group, treatment with sulfur trioxidepyridine complex, and O-deacylation, into the title sulfo Lex analog 22. Compound 23 was transformed by reductive removal of the benzyl groups, O-acetylation, Odeacylation and subsequent hydrolysis of the methyl ester, into the sLe<sup>x</sup> epitope analog 25.

Copyright © 1995 by Marcel Dekker, Inc.

## **INTRODUCTION**

The selectins<sup>1-4</sup> [E-selectin (ELAM-1), L-selectin (LECAM-1) and P-selectin (PADGEM)] are a family of cell-adhesion receptors and recognize<sup>5-9</sup> the sialyl Le<sup>x</sup> and sialyl Le<sup>a</sup> determinants, which are found as the terminal carbohydrate structures of both cell membrane glycolipids and glycoproteins, and related oligosaccharides.<sup>11,12</sup> Binding of selectins to their carbohydrate ligands appears to be required for neutrophil extravasation, and plays a major role in lymphocyte recirculation and platelet adhesion. Based on the three-dimensional structure<sup>9</sup> of sLe<sup>x</sup> and sLe<sup>a</sup>, various types of sLe<sup>x</sup> analogs<sup>13-17</sup> containing chemically modified sialic acid, fucose, galactose, and *N*-acetylglucosamine moieties have been synthesized in order to clarify the structural features of each component of the carbohydrate ligand required for selectin recognition.<sup>18-19</sup>

Recently, it has been demonstrated<sup>19</sup> that E- and L-selectin can bind to sulfo-Le<sup>x</sup>-like structures isolated from an ovarian cyst adenoma, and using chemically synthesized sulfo Le<sup>a</sup> and sulfo Le<sup>x</sup> oligosaccharides, sulfo Le<sup>a</sup> pentasaccharide has the most potent activity for E-selectin recognition. As a part of our continuing studies on structure-activity correlations in the sLe<sup>x</sup> epitope, we describe the synthesis of sLe<sup>x</sup> and sulfo Le<sup>x</sup> epitope analogs containing the 1-deoxy-N-acetylglucosamine in place of *N*-acetylglucosamine moiety.

## **RESULTS AND DISCUSSION**

For the synthesis of the desired sulfo and sialyl Le<sup>x</sup> epitope analogs we employed 2,4,6-tri-*O*-benzoyl-3-levulinyl- $\alpha$ -D-galactopyranosyl trichloroacetimidate (6) as the glycosyl donor and 2-acetamido-1,5-anhydro-3,6-di-*O*-benzyl-2-deoxy-D-glucitol (10) as a suitably protected glycosyl acceptor. The acceptor 10 was coupled with 6 using boron trifluoride etherate,<sup>20</sup> or with the methyl thioglycoside derivative<sup>21</sup> (11) of  $\alpha$ -sialyl-(2 $\rightarrow$ 3)-galactose as the donor using dimethyl(methylthio)sulfonium triflate<sup>22</sup> (DMTST), as promoter, to afford the corresponding di- and trisaccharides 12 and 15. The di- and trisaccharide acceptors were then glycosylated with methyl 2,3,4-tri-*O*-

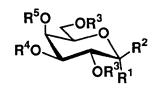
benzyl-1-thio- $\beta$ -L-fucopyranoside<sup>23</sup> (17). By further processing according to our usual procedure<sup>24</sup> the resulting tri- and tetrasaccharide intermediates 18 and 23 could be converted into the end products 22 and 25.

Treatment of 2-(trimethylsilyl)ethyl  $\beta$ -D-galactopyranoside<sup>25</sup> (1) with 2,2dimethoxypropane in *N*,*N*-dimethylformamide (DMF) in the presence of *p*toluenesulfonic acid monohydrate at 80 °C gave the 3,4-*O*-isopropylidene derivative, and this on *O*-benzoylation gave 2 in 77% yield. Hydrolysis of the isopropylidene group in 2 with aq 80% acetic acid gave the 2,6-di-*O*-benzoyl derivative 3. Treatment of 3 with levulinic anhydride in pyridine for one h at -50 °C gave the 3-*O*-levulinyl derivative 4 in 87% yield, exclusively, and this on *O*-benzoylation and subsequent hydrolysis<sup>25</sup> of the 2-(trimethylsilyl)ethyl group with trifluoroacetic acid followed by treatment<sup>20,26</sup> with trichloroacetonitrile in the presence of 1,8diazabicyclo[5.4.0]undec-7-ene (DBU), was converted into the glycosyl donor 6 in good yield. The <sup>1</sup>H NMR data of 6 [ $\delta$  6.86 (J<sub>1,2</sub> = 3.1 Hz, H-1), 8.62 (C = NH)] established the anomeric configuration of the imidate.

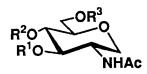
Treatment of 2-acetamido-1,2-anhydro-2-deoxy-D-glucitol<sup>27</sup> (7) with benzaldehyde dimethyl acetal in DMF in the presence of *p*-toluenesulfonic acid monohydrate gave the 4,6-O-benzylidene derivative **8** (97%). Treatment of **8** with benzyl bromide in DMF in the presence of sodium hydride for 3 h at room temperature gave the 3-O-benzyl derivative (71%) as crystals which, on reductive ring-opening<sup>28</sup> of the benzylidene group with sodium cyanoborohydride-hydrogen chloride in tetrahydrofuran, afforded the desired glycosyl acceptor **10** (91%) as crystals.

Glycosylation<sup>20</sup> of 10 with 1.5 equiv of 6 in dichloromethane for 3 h at room temperature in the presence of boron trifluoride etherate and 4Å molecular sieves gave the expected disaccharide derivative 12 (58%) which, on catalytic hydrogenolysis (10% Pd-C) in ethanol-acetic acid of the benzyl group and subsequent 6-O-benzoylation, gave the next glycosyl acceptor 14 in good yield.

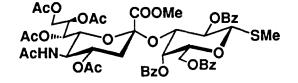
On the other hand, glycosylation<sup>21</sup> of **10** with 1.7 equiv of **11** in dichloromethane in the presence of DMTST and 4Å molecular sieves for 48 h at room temperature gave the trisaccharide **15** (92%) which, on reductive removal of the benzyl groups and subsequent 6-O-benzoylation as described for the preparation of **14**, gave another glycosyl acceptor **16** (76%).



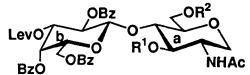
	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R⁴	R <sup>5</sup>
1	Н	OSE	Н	Н	Н
2	н	OSE	Bz	ip	d
3	н	OSE	Bz	Н	н
4	н	OSE	Bz	Lev	Н
5	Н	OSE	Bz	Lev	Bz
6	OC(=NH)CCl <sub>3</sub>	Н	Bz	Lev	Bz

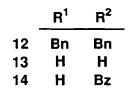


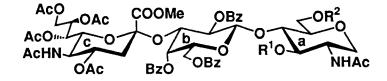
	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>
7	Н	Н	Н
8	Н	benzy	lidene
9	Bn	benzy	lidene
10	Bn	н	Bn

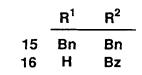


11













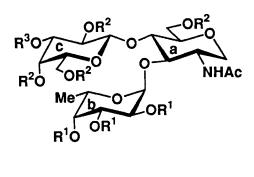
Glycosylation of 14 or 16 with methyl 2,3,4-tri-O-benzyl-1-thio- $\beta$ -Lfucopyranoside (17) in benzene in the presence of DMTST and 4Å molecular sieves afforded the corresponding  $\alpha$ -glycosides 18 (76%) and 23 (88%), respectively. Hydrogenolysis of the benzyl group in 18 followed by acetylation gave 19 (81%). Significant signals in the <sup>1</sup>H NMR spectrum of 19 were at  $\delta$  4.78 (d, J<sub>1,2</sub> = 8.1 Hz, H-1c) and 5.43 (d, J<sub>1,2</sub> = 3.8 Hz, H-1b), which showed the anomeric configurations to be  $\beta$  and  $\alpha$  respectively.

Treatment of 19 with hydrazine monoacetate in ethanol for 3 h at room temperature gave O(2,4,6-tri-O-benzoyl- $\beta$ -D-galactopyranosyl)- $(1\rightarrow 4)$ -O[(2,3,6-tri-O-acetyl- $\alpha$ -L-fucopyranosyl)- $(1\rightarrow 3)$ ]-2-acetamido-1,5-anhydro-6-O-benzoyl-D-glucitol (20) in 66% yield. When treated with sulfur trioxide pyridine complex in DMF for one h at room temperature, compound 20 gave the sulfated Le<sup>x</sup> analog derivative 21 (93%), and this was transformed by removal of the protecting groups into O(3-Osulfo- $\beta$ -D-galactopyranosyl)- $(1\rightarrow 4)$ - $O[\alpha$ -L-fucopyranosyl- $(1\rightarrow 3)$ ]-2-acetamido-1,5anhydro-D-glucitol sodium salt (22) quantitatively.

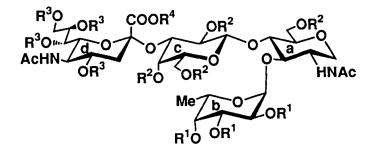
Hydrogenolysis of the benzyl groups in 23 and subsequent acetylation gave the protected sLe<sup>x</sup> analog 24 in 85% yield. Significant signals in the <sup>1</sup>H NMR spectrum of 24 were at  $\delta$  5.35 (d, J<sub>1,2</sub> = 3.8 Hz, H-1b) and 5.48 (dd, J<sub>1,2</sub> = 8.1 Hz, J<sub>2,3</sub> = 10.1 Hz, H-2c), indicating the expected anomeric configurations. Finally, *O*-deacylation of 24 with sodium methoxide in methanol and hydrolysis of the methyl ester group yielded the desired sLe<sup>x</sup> epitope analog 25 in almost quantitative yield after chromatography on a column of Sephadex LH-20.

#### **EXPERIMENTAL**

General Procedures. Specific rotations were determined with a Union PM-201 polarimeter at 25 °C, and IR spectra were recorded with a Jasco A-100 spectrophotometer. <sup>1</sup>H NMR spectra were recorded with a JEOL JNM-GX 270 spectrometer. Mass spectra were recorded using an electroscopy mass spectrometer (Perkin-Elmer Sciex Instrument, Thornhill, Canada) fitted with an atmospheric pressure ionization source. Preparative chromatography was performed on silica gel



_	R <sup>1</sup>	$R^2$	R <sup>3</sup>
18	Bn	Bz	Lev
19	Ac	Bz	Lev
20	Ac	Bz	н
21	Ac	Bz	SO₃ • Pyr
22	Н	Н	SO <sub>3</sub> Na



	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>	
23	Bn	Bz	Ac	Ме	
24	Ac	Bz	Ac	Ме	SE = 2-(trimethylsilyl)ethyl
25	Н	Н	Н	Н	Bz = benzoyl
					Bn = benzyl
					Lev = CH <sub>3</sub> COCH <sub>2</sub> CH <sub>2</sub> CO
					ipd = isopropylidene

(Wako Chemical Co., 200 mesh) with the solvent systems specified. Concentrations were conducted *in vacuo*.

2-(Trimethylsilyl)ethyl 2,6-Di-O-benzoyl-3,4-O-isopropylidene- $\beta$ -D-galactopyranoside (2). To a solution of 2-(trimethylsilyl)ethyl  $\beta$ -D-galactopyranoside<sup>25</sup> (1, 2.7 g, 9.4 mmol) in N,N-dimethylformamide (DMF, 10 mL) were added 2,2-dimethoxypropane (2.3 mL) and p-toluenesulfonic acid monohydrate (50 mg) and the mixture was stirred for 2 h at 80 °C; the course of the reaction was

monitored by TLC. After completion of the reaction, the solution was neutralized with Na<sub>2</sub>CO<sub>3</sub> and filtered. The solution was concentrated to a syrup, which was treated with benzoyl chloride (2.1 mL) in pyridine (4 mL) for 30 min at room temperature. MeOH (1 mL) was added to the mixture and concentrated to a syrup, which was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The extract was successively washed with 2 M HCl, M Na<sub>2</sub>CO<sub>3</sub> and water, dried (Na<sub>2</sub>SO<sub>4</sub>), and concentrated. Column chromatography (1:10 EtOAc-hexane) of the residue on silica gel (150 g) gave **2** (3.8 g, 77%) as crystals. Recrystallization from EtOAc-hexane gave needles: mp 123 °C;  $[\alpha]_D$  +19.0° (*c* 1.7, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.00 (m, 2H, Me<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>), 1.47, 1.75 (2s, 6H, Me<sub>2</sub>C), 3.66, 4.06 (2m, 2H, Me<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>), 4.31 (m, 1H, H-5), 4.48 (dd, 1H, J<sub>2,3</sub> = 7.2 Hz, J<sub>3,4</sub> = 4.5 Hz, H-3), 4.65 (d, 1H, J<sub>1,2</sub> = 8.3 Hz, H-1), 4.78 (m, 2H, H-6, 6'), 5.36 (dd, 1H, H-2), and 7.37-8.19 (m, 10H, 2Ph).

Anal. Calcd for C<sub>23</sub>H<sub>36</sub>O<sub>8</sub>Si (528.7): C, 52.25; H, 6.86. Found: C, 52.30; H, 6.85.

**2-(Trimethylsilyl)ethyl 2,6-Di-***O***-benzoyl**-β**-D-galactopyranoside** (3). A solution of **2** (3.8 g, 6.4 mmol) in aq 80% AcOH (50 mL) was heated for 4 h at 60 °C then concentrated. Column chromatography (100:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the residue on silica gel (150 g) gave **3** (3.1 g, quantitative) as crystals. Recrystallization from EtOAc-hexane gave needles: mp 124 °C;  $[\alpha]_D$  -15.0° (*c* 1.1, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.00 (m, 2H, Me<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>), 3.68, 4.04 (2m, 2H, Me<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>), 4.04 (m, 2H, H-5, H-6), 4.12 (d, 1H, J<sub>3</sub>,4 = 4.0 Hz, H-4), 4.69 (d, 1H, J<sub>1,2</sub> = 7.7 Hz, H-1), 4.70 (dd, 1H, J<sub>2,3</sub> = 9.5 Hz, H-3), 4.76 (dd, 1H, J<sub>gem</sub> = 11.3 Hz, J<sub>5,6</sub>' = 6.2 Hz, H-6'), 5.33 (dd, 1H, H-2), and 7.35-8.16 (m, 10H, 2Ph).

Anal. Calcd for C<sub>25</sub>H<sub>32</sub>O<sub>8</sub>Si (488.6): C, 61.45; H, 6.60. Found: C, 61.43; H, 6.67.

2-(Trimethylsilyl)ethyl 2,6-Di-O-benzoyl-3-O-levulinyl- $\beta$ -D-galactopyranoside (4). To a solution of 3 (1.0 g, 2 mmol) in pyridine (10 mL), cooled to -50 °C, were added levulinic anhydride (780 mg, 4 mmol), and the mixture was stirred for 10 min at -50 °C. After completion of the reaction, MeOH (0.5 mL) was added and the mixture was concentrated then extracted with CH<sub>2</sub>Cl<sub>2</sub>. The extract was washed with 2M HCl and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography (200:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the residue on silica gel (70 g) gave 4 (1.04 g, 87%) as an amorphous mass:  $[\alpha]_D$  +4.2° (c 1.9, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.0 (m, 2H, Me<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>), 2.52-2.82 (m, 4H, MeCOCH<sub>2</sub>CH<sub>2</sub>), 4.36 (br d, 1H, J<sub>3,4</sub> = 3.3 Hz, H-4), 4.75 (d, 1H, J<sub>1,2</sub> = 7.9 Hz, H-1), 5.18 (dd, 1H, J<sub>2,3</sub> = 10.3 Hz, H-3), 5.68 (dd, 1H, H-2), and 7.34-8.18 (m, 10H, 2Ph).

Anal. Calcd for C30H38O10Si (586.7): C, 61.41; H, 6.53. Found: C, 61.38; H, 6.45.

2-(Trimethylsilyl)ethyl 2,4,6-Tri-*O*-benzoyl-3-*O*-levulinyl-β-Dgalactopyranoside (5). To a solution of 4 (2.27 g, 3.85 mmol) in pyridine (1 mL) and CH<sub>2</sub>Cl<sub>2</sub> (10 mL), cooled to 0 °C, was added benzoyl chloride (0.7 mL, 4.8 mmol) and the mixture was stirred overnight at room temperature. Processing as described for 4 gave 5 (2.44 g, 89%) as an amorphous mass:  $[\alpha]_D$  +23.3° (*c* 0.9, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.0 (m, 2H, Me<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>), 2.38-2.78 (m, 4H, MeCOCH<sub>2</sub>CH<sub>2</sub>), 3.70, 4.11 (2m, 2H, Me<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>), 4.30 (m, 1H, H-5), 4.46 (dd, 1H, J<sub>gem</sub> = 11.1 Hz, J<sub>5,6</sub> = 6.9 Hz, J<sub>5,6</sub>' = 6.4 Hz, H-6'), 4.71 (dd, 1H, H-6), 4.82 (d, 1H, J<sub>1,2</sub> = 7.9 Hz, H-1), 5.43 (dd, 1H, J<sub>2,3</sub> = 10.4 Hz, J<sub>3,4</sub> = 3.6 Hz, H-3), 5.67 (dd, 1H, H-2), 5.90 (d, 1H, H-4), and 7.33-8.25 (m, 15H, 3Ph).

Anal. Calcd for C37H42O11Si (690.8): C, 64.33; H, 6.13. Found: C, 64.53; H, 6.03.

2,4,6-Tri-O-benzoyl-3-O-levulinyl- $\alpha$ -D-galactopyranosyl Trichloroacetimidate (6). A solution of 5 (2.27 g, 3.2 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) and CF<sub>3</sub>CO<sub>2</sub>H (5 mL) was stirred overnight at room temperature then concentrated. Column chromatography (1:2 EtOAc-hexane) of the residue on silica gel (100 g) gave the 1-hydroxy compound (2.05 g, quantitative). Drierite (250 mg) was added to a solution of the 1-hydyoxy compound (500 mg, 0.85 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (1 mL), which was then cooled to -5 °C. Cl<sub>3</sub>CCN (0.26 mL) and 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU, 0.07 mL) were then added and the mixture was stirred for one h at room temperature and then concentrated. Column chromatography (1:4 EtOAc-hexane) of the residue on silica gel (50 g) gave 6 (520 mg, 83%) as an amorphous mass:  $[\alpha]_D$  +84.5° (*c* 0.3, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  2.33-2.73 (m, 4H, MeCOCH<sub>2</sub>CH<sub>2</sub>), 4.04 (dd, 1H, J<sub>gem</sub> = 11.4 Hz, J<sub>5</sub>,6 = 6.6 Hz, J<sub>5</sub>,6' = 5.9 Hz, H-6'), 4.57 (dd, 1H, H-6), 4.77 (m, 1H, H-5), 5.76 (dd, 1H, J<sub>2</sub>,3 = 10.8 Hz, J<sub>3</sub>,4 = 3.3 Hz, H-3), 5.82 (dd, 1H,  $J_{1,2}$ = 3.1 Hz, H-2), 6.00 (br d, 1H, H-4), 6.86 (d, 1H, H-1), 7.26-8.15 (m, 15H, 3Ph), and 8.62 (s, 1H, C = NH).

Anal. Calcd for C34H30Cl3NO11 (735.0): C, 55.56; H, 4.11; N, 1.91. Found: C, 55.38; H, 4.10; N, 1.65.

2-Acetamido-1,5-anhydro-4,6-O-benzylidene-2-deoxy-D-glucitol (8). To a solution of  $7^{27}$  (4.97 g, 24.2 mmol) in DMF (50 mL) was added Drierite (5 g) and the mixture was stirred for 3 h at room temperature. Benzaldehyde dimethyl acetal (7.3 mL) and p-toluenesulfonic acid monohydrate (50 mg) were added to the mixture which was stirred overnight at room temperature and then neutralized with NaHCO3. The precipitate was filtered off and the solution was concentrated. Column chromatography (50:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the residue on silica gel (400 g) gave 8 (6.9 g, 97%) as crystals. Recrystallization from ether-hexane gave needles: mp 196 °C;  $[\alpha]_D$  -41.7° (c 1.1, MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.99 (s, 3H, AcN), 3.23 (t, 1H, Jgem = J<sub>1</sub>ax, 2 = 9.3 Hz, H-1ax), 3.38 (m, 1H, H-5), 3.53 (t, 1H, J<sub>3</sub>, 4 = J<sub>4</sub>, 5 = 9.2 Hz, H-4), 3.69 (t, 1H, J<sub>2</sub>, 3 = J<sub>3</sub>, 4 = 9.3 Hz, H-3), 3.73 (t, 1H, Jgem = J<sub>5</sub>, 6ax = 10.4 Hz, H-6ax), 3.93 (dd, 1H, J<sub>1</sub>eq, 2 = 5.5 Hz, H-1eq), 4.00 (m, 1H, H-2), 4.28 (dd, 1H, J<sub>gem</sub> = 10.4 Hz, J<sub>5</sub>, 6eq = 4.9 Hz, H-6eq), 4.60 (d, 1H, NH), 5.58 (s, 1H, CH-Ph), and 7.34-7.55 (m, 5H, Ph).

Anal. Calcd for C<sub>15</sub>H<sub>1</sub>9NO<sub>5</sub> (293.3): C, 61.42; H, 6.53; N, 4.78. Found: C, 61.30; H, 6.41; N, 4.79.

2-Acetamido-1,5-anhydro-3-O-benzyl-4,6-O-benzylidene-2-deoxy-D-glucitol (9). To a solution of 8 (3.5 g, 11.9 mmol) in DMF (40 mL) was added a suspension of sodium hydride in oil (430 mg, 60% of sodium hydride by weight). The mixture was stirred for one h at 0 °C, benzyl bromide (2.13 g) was added dropwise, and stirring was continued for 3 h at room temperature. Methanol (1 mL) was added and the mixture was concentrated then extracted with CH<sub>2</sub>Cl<sub>2</sub>. The extract was washed with water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Recrystallization from etherhexane gave 9 (3.25 g, 71%) as needles: mp 269 °C;  $[\alpha]_D$  -43.2° (*c* 1.1, 1:1 CHCl<sub>3</sub>-MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.88 (s, 3H, AcN), 3.21 (t, 1H, J<sub>gem</sub> = J<sub>1ax,2</sub> = 12.7 Hz, H-1ax), 3.38 (m, 1H, H-5), 3.93 (dd, 1H, J<sub>1eq,2</sub> = 5.3 Hz, H-1eq), 4.31 (dd, 1H, J5,6eq = 4.8 Hz, J<sub>gem</sub> = 10.4 Hz, H-6eq), 5.60 (s, 1H, CH-Ph), and 7.27-7.56 (m, 10H, 2Ph). Anal. Calcd for C<sub>22</sub>H<sub>25</sub>NO<sub>5</sub> (383.4): C, 68.91; H, 6.57; N, 3.65. Found: C, 68.73; H, 6.61; N, 3.58.

2-Acetamido-1,5-anhydro-3,6-di-O-benzyl-2-deoxy-D-glucitol (10). To a solution of 9 (3.6 g, 9.4 mmol) in dry THF (36 mL) were added powdered MS-3Å (7.2 g), and the mixture was stirred for one h at room temperature, and sodium cyanoborohydride (8.8 g) was gradually added. After the reagent had dissolved, hydrogen chloride in ether was added dropwise at room temperature until the evolution of gas ceased. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (100 mL), and this was washed with water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography (1:1 EtOAc-hexane) of the residue on silica gel (150 g) gave **10** (3.3 g, 91%) as crystals. Recrystallization from ether-hexane gave needles: mp 152 °C;  $[\alpha]_D$  +18.5° (*c* 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.86 (s, 3H, AcN), 6.18 (br s, 1H, OH), 7.25-7.56 (m, 10H, 2Ph).

Anal. Calcd for C22H27NO5 (385.4): C, 68.55; H, 7.06; N, 3.63. Found: C, 68.51; H, 7.20; N, 3.50.

*O* - (2,4,6 - Tri - *O* - benzoyl - 3 - *O* - levulinyl - β - D - galactopyranosyl)-(1→4)-2 - acetamido - 1,5 - anhydro - 3,6 - di - *O* - benzyl - 2 - deoxy - D - glucitol (12). To a solution of 6 (1.2 g, 1.63 mmol) and 10 (420 mg, 1.1 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (6 mL) were added MS-4Å (AW-300, 2 g), and the mixture was stirred overnight at room temperature, then cooled to 0 °C. Boron trifluoride etherate (20 µL) was added and this was stirred for 3 h at room temperature. After completion of the reaction, the mixture was neutralized with Et<sub>3</sub>N and filtered. The precipitate was washed with CH<sub>2</sub>Cl<sub>2</sub>. The combined filtrate and washings were successively washed with M Na<sub>2</sub>CO<sub>3</sub> and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography (100:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the residue on silica gel (100 g) gave 12 (595 mg, 58%) as an amorphous mass: [α]<sub>D</sub> +11.1° (*c* 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.88, 1.91 (2s, 6H, Ac, AcN), 2.38, 2.55 (2m, 4H, MeCOCH<sub>2</sub>CH<sub>2</sub>), 3.07 (t, 1H, J<sub>gem</sub> = J<sub>1</sub>ax, 2 = 8.1 Hz, H-1aax), 4.75 (4d, 4H, 2CH<sub>2</sub>Ph), 5.25 (dd, 1H, J<sub>2</sub>, 3 = 10.6 Hz, J<sub>3</sub>, 4 = 3.4 Hz, H-3b), 5.56 (dd, 1H, J<sub>1,2</sub> = 8.2 Hz, H-2b), 5.75 (d, 1H, H-4b), and 7.23-8.13 (m, 25H, 5Ph).

Anal. Calcd for C54H55NO15 (958.0): C, 67.70; H, 5.79; N, 1.46. Found: C, 67.53; H, 5.88; N, 1.50.

O - (2,4,6-Tri-O-benzoyl-3 - O-levulinyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-2-acetamido-1,5-anhydro-2-deoxy-glucitol (13). A solution of 12 (593 mg, 0.63 mmol) in EtOH (30 mL) and AcOH (7 mL) was hydrogenolyzed in the presence of 10% Pd-C (600 mg) overnight at 40 °C, then filtered, and concentrated. Column chromatography (80:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the residue on silica gel (60 g) gave 13 (433 mg, 90%) as an amorphous mass:  $[\alpha]_D$  +63.5° (*c* 1.2, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.88, 1.91 (2s, 6H, Ac, AcN), 2.38, 2.55 (2m, 4H, MeCOCH<sub>2</sub>CH<sub>2</sub>), 3.07 (t, 1H, J<sub>gem</sub> = J<sub>1</sub>*ax*,2 = 8.1 Hz, H-1a*ax*), 5.25 (dd, 1H, J<sub>2,3</sub> = 10.6 Hz, J<sub>3,4</sub> = 3.4 Hz, H-3b), 5.56 (dd, 1H, J<sub>1,2</sub> = 8.2 Hz, J<sub>2,3</sub> = 10.2 Hz, H-2b), 5.67 (d, 1H, J<sub>2,NH</sub> = 7.1 Hz, NH), 5.75 (d, 1H, H-4b), and 7.23-8.13 (m, 15H, 3Ph).

Anal. Calcd for C40H43NO15 (777.8): C, 61.76; H, 5.57; N, 1.80. Found: C, 61.77; H, 5.43; N, 1.62.

*O* - (2,4,6 - Tri - *O* - benzoyl - 3 - *O* - levulinyl - β - D - galactopyranosyl) - (1→4)-2-acetamido-1,5-anhydro-6-*O*-benzoyl-2-deoxy-D-glucitol (14). To a solution of 13 (62 mg, 0.08 mmol) in pyridine (1 mL) and CH<sub>2</sub>Cl<sub>2</sub> (1 mL), cooled to -40 °C, was added benzoyl chloride (0.11 mL), and the mixture was stirred for one h at -40 °C. MeOH (0.1 mL) was added, and the mixture was concentrated then extracted with CH<sub>2</sub>Cl<sub>2</sub>. The extract was washed with 2 M HCl, M Na<sub>2</sub>CO<sub>3</sub> and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography (1:2 EtOAchexane) of the residue on silica gel (20 g) gave 14 (67 mg, 95%) as an amorphous mass:  $[\alpha]_D$  +84.5° (*c* 1.3, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.87, 1.91 (2s, 6H, Ac, AcN), 2.34, 2.48 (2m, 4H, MeCOCH<sub>2</sub>CH<sub>2</sub>), 3.08 (t, 1H, J<sub>gem</sub> = J<sub>1ax,2</sub> = 10.7 Hz, H-1aax), 4.95 (d, 1H, J<sub>1,2</sub> = 8.2 Hz, H-1b), 5.38 (dd, 1H, J<sub>2,3</sub> = 10.5 Hz, J<sub>3,4</sub> = 3.2 Hz, H-3b), 5.72 (dd, 1H, H-2b), 5.75 (d, 1H, J<sub>2,NH</sub> = 7.1 Hz, NH), 5.83 (d, 1H, H-4b), and 7.15-8.14 (m, 20H, 4Ph).

Anal. Calcd for C47H47NO16 (881.9): C, 64.01; H, 5.37; N, 1.59. Found: C, 64.23; H, 5.40; N, 1.56.

*O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- $\alpha$ -D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-benzoyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 4)-2-acetamido-1,5-anhydro-3,6-di-*O*benzyl-2-deoxy-D-glucitol (15). To a solution of 11<sup>21</sup> (220 mg, 0.22 mmol) and 10 (52 mg, 0.13 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (1.5 mL) were added MS-4Å (250 mg) and the mixture was stirred overnight at room temperature then cooled to 0 °C. Dimethyl(methylthio)sulfonium triflate (DMTST, 260 mg) and MS-4Å (260 mg) were added to the mixture, and this was stirred for 2 days at room temperature. Reaction progress was monitored by TLC. The precipitate was collected and washed with CH<sub>2</sub>Cl<sub>2</sub>, and the combined filtrate and washings were washed with M NaHCO3 and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography (3:1 EtOAchexane) of the residue on silica gel (30 g) gave 15 (165 mg, 92%) as an amorphous mass:  $[\alpha]_D$  +24.3° (*c* 0.4, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.61-2.15 (6s, 18H, 4AcO, 2AcN), 2.46 (dd, 1H, J<sub>gem</sub> = 12.5 Hz, J<sub>3eq</sub>, 4 = 4.5 Hz, H-3ceq), 3.76 (s, 3H, MeO), 4.50 (4d, 4H, 2CH<sub>2</sub>Ph), 4.83 (m, 1H, J<sub>3ax</sub>, 4 = J4, 5 = 11.2 Hz, H-4c), 5.04 (d, 1H, J<sub>1,2</sub> = 7.9 Hz, H-1b), 5.40 (d, 1H, H-4b), 5.50 (dd, 1H, J<sub>2,3</sub> = 10.1 Hz, H-2b), 5.68 (m, 1H, H-8c), and 7.21-8.20 (m, 25H, 5Ph).

Anal. Calcd for C69H75N2O24 (1333.4): C, 62.15; H, 5.67; N, 2.10. Found: C, 62.03; H, 5.77; N, 1.95.

*O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-*glyc-ero*-α-D-*galacto*-2-nonulopyranosylonate) - (2→3) - *O* - (2,4,6-tri-*O*-benzoyl-2-deoxy-D-glucitol (16). A solution of 15 (462 mg, 0.35 mmol) in EtOH (40 mL) and AcOH (7 mL) was stirred with 10% Pd-C (300 mg) overnight at 45 °C under hydrogen. The catalyst was collected and washed with MeOH, the combined filtrate and washings was concentrated. To a solution of the residue in pyridine (0.5 mL) and CH<sub>2</sub>Cl<sub>2</sub> (1.5 mL), cooled to -50 °C, was added a solution of benzoyl chloride (0.38 mL) in CH<sub>2</sub>Cl<sub>2</sub> (4 mL), and the mixture was stirred for 3 h at -50~-20 °C; the course of the reaction was monitored by TLC. A workup as described for 14 gave 16 (146 mg, 84%) as an amorphous mass: [α]<sub>D</sub> +51.0° (*c* 1.7, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.48-2.11 (6s, 18H, 4AcO, 2AcN), 2.43 (dd, 1H, Jgem = 12.6 Hz, J<sub>3</sub>eq, 4 = 4.5 Hz, H-3ceq), 3.02 (t, 1H, Jgem = J<sub>1</sub>ax, 2 = 10.8 Hz, H-1aax), 4.78 (m, 1H, H-4c), 5.09 (d, 1H, J<sub>1</sub>, 2 = 7.9 Hz, H-1b), 5.23 (dd, 1H, H-2b), and 7.15-8.13 (m, 20H, 4Ph).

Anal. Calcd for C<sub>62</sub>H<sub>67</sub>N<sub>2</sub>O<sub>25</sub> (1257.2): C, 59.23; H, 5.37; N, 2.23. Found: C, 59.41; H, 5.44; N, 2.25.

*O* - (2,4,6 - Tri - *O*-benzoyl - 3 - *O* - levulinyl - β - D - galactopyranosyl) - (1→4) - *O* - [(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-(1→3)] - 2-acetamido-1,5-anhydro-6-*O*-benzoyl-2-deoxy-D-glucitol (18). To a solution of 14 (68 mg, 0.078 mmol) and methyl 2,3,4-tri-*O*-benzyl-1-thio-β-L-fucopyranoside<sup>23</sup> (17, 43 mg, 0.092 mmol) in benzene (0.5 mL) were added MS-4Å (400 mg), and the mixture was stirred for 4 h at room temperature then cooled to 0 °C. DMTST (80 mg) was added to the mixture, and this was stirred overnight at 5 °C. The precipitate was collected and washed with CH<sub>2</sub>Cl<sub>2</sub>. The combined filtrate and washings were concentrated to a syrup which was chromatographed on a column of silica gel (20 g) with 100:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH to give 18 (77 mg, 76%) as an amorphous mass: [α]<sub>D</sub> -16.0° (*c* 1.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.05 (d, 3H, J<sub>5,6</sub> = 6.2 Hz, H-6b), 1.89 (s, 3H, AcN), 2.04 (s, 3H, Ac), 2.04-2.55 (m, 4H, MeCOCH<sub>2</sub>CH<sub>2</sub>), 3.08 (t, 1H, H-1aax), 4.95 (d, 1H, J<sub>1,2</sub> = 10.3 Hz, H-1c), 5.42 (dd, 1H, H-3c), 5.57 (dd, 1H, J<sub>2,3</sub> = 10.5 Hz, H-2c), 5.83 (d, 1H, H-4c), and 7.23-8.17 (m, 35H, 7Ph).

Anal. Calcd for C74H75NO<sub>20</sub> (1298.4): C, 68.45; H, 5.82; N, 1.08. Found: C, 68.31; H, 5.85; N, 1.03.

 $O \cdot (2,4,6 \cdot \text{Tri} \cdot O \cdot \text{benzoyl} \cdot 3 \cdot O \cdot \text{levulinyl} \cdot \beta \cdot D \cdot \text{galactopyranosyl}) \cdot (1 \rightarrow 4) \cdot O \cdot [(2,3,4 \cdot \text{tri} \cdot O \cdot \text{acetyl} \cdot \alpha \cdot \text{L-fucopyranosyl}) \cdot (1 \rightarrow 3)] \cdot 2 \cdot \text{acetamido} \cdot 1,5 \cdot \text{anhydro} \cdot 6 \cdot O \cdot \text{benzoyl} \cdot 2 \cdot \text{deoxy} \cdot D \cdot \text{glucitol}$  (19). A solution of 18 (75 mg, 0.058 mmol) in EtOH (4 mL) and AcOH (1 mL) was stirred with 10% Pd-C (75 mg) for 12 h at 40 °C under hydrogen. The catalyst was collected and the solution was concentrated, the residue was treated with Ac2O (0.5 mL) and pyridine (1 mL) overnight at room temperature. The mixture was concentrated then extracted with CH<sub>2</sub>Cl<sub>2</sub>, and the extract was washed with 2 M HCl and M Na<sub>2</sub>CO<sub>3</sub>, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography (100:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the residue on silica gel (10 g) gave 19 (54 mg, 81%) as an amorphous mass:  $[\alpha]_D \cdot 14.5^\circ$  (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.18 (d, 3H, J<sub>5,6</sub> = 6.4 Hz, H-6b), 1.91-2.16 (5s, 15H, 3AcO, Ac, AcN), 3.17 (t, 1H, H-1aax), 4.78 (d, 1H, J<sub>1,2</sub> = 8.1 Hz, H-1c), 5.15 (dd, 1H, J<sub>2,3</sub> = 8.3 Hz, J<sub>3,4</sub> = 3.3 Hz, H-3c), 5.43 (d, 1H, J<sub>1,2</sub> = 3.8 Hz, H-1b), 5.53 (t, 1H, H-2c), 5.77 (d, 1H, H-4c), and 7.27-8.11 (m, 20H, 4Ph).

Anal. Calcd for C59H63NO23 (1154.2): C, 61.39; H, 5.50; N, 1.21. Found: C, 61.48; H, 5.68; N, 1.12.

 $O \cdot (2,4,6$ -Tri-O-benzoyl- $\beta$ -D-galactopyranosyl)- $(1 \rightarrow 4)$ - $O \cdot [(2,3,6$ -tri-O-acetyl- $\alpha$ -L-fucopyranosyl)- $(1 \rightarrow 3)$ ]-2-acetamido-1,5-anhydro-6-O-benzoyl-D-glucitol (20). To a solution of 19 (54 mg, 0.047 mmol) in EtOH (1 mL) was added hydrazine monoacetate (5.2 mg), and the mixture was stirred for 3 h at room temperature then concentrated. Column chromatography (80:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the residue on silica gel (10 g) gave 20 (33 mg, 66%) as an amorphous mass:  $[\alpha]_D$ -31.1° (c 1.7, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.18 (d, 3H, J<sub>5,6</sub> = 6.4 Hz, H-6b), 1.87-2.10 (4s, 12H, 3AcO, AcN), 3.07 (t, 1H, H-1aax), 4.13 (dd, 1H, J<sub>2,3</sub> = 10.0 Hz, J<sub>3,4</sub> = 3.3 Hz, H-3c), 5.13 (dd, 1H, J<sub>2,3</sub> = 10.8 Hz, J<sub>3,4</sub> = 3.9 Hz, H-3b), 5.63 (d, 1H, H-4c), 6.16 (d, 1H, J<sub>NH,2</sub> = 9.3 Hz, NH), and 7.27-8.02 (m, 20H, 4Ph).

Anal. Calcd for C54H57NO21 (1056.0): C, 61.41; H, 5.44; N, 1.33. Found: C, 61.35; H, 5.64; N, 1.31.

O - (2,4,6-Tri-O-benzoyl-3-sulfo- $\beta$ -D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[(2,3,4-tri-O-acetyl- $\alpha$ -L-fucopyranosyl)- $(1\rightarrow 3)$ ]-2-acetamido-1,5-anhydro-6-O-benzoyl-D-glucitol pyridine salt (21). To a solution of 20 (33 mg, 0.031 mmol) in DMF (0.3 mL) was added sulfur trioxide pyridine complex (25 mg), and the mixture was stirred for one h. Methanol (0.1 mL) was added to the mixture and concentrated at 25 °C. Column chromatography (15:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the residue on silica gel (10 g) gave 21 (35 mg, 93%) as an amorphous mass:  $[\alpha]_D$ -5.5° (c1.2, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.01 (d, 3H, J<sub>5,6</sub> = 6.2 Hz, H-6b), 1.88-2.08 (4s, 12H, 3AcO, AcN), and 7.10-8.10 (m, 25H, 4Ph, C5H<sub>5</sub>N).

Anal. Calcd for C59H62N2O24S (1215.2): C, 58.31; H, 5.14; N, 2.31. Found: C, 58.58; H, 5.35; N, 2.20.

*O*-(3-*O*-Sulfo-β-D-galactopyranosyl)-(1→4)-*O*-[α-L-fucopyranosyl-(1→3)]-2-acetamido-1,5-anhydro-D-glucitol sodium salt (22). To a solution of 21 (50 mg, 0.041 mmol) in MeOH (1 mL) was added NaOMe (5 mg) and the mixture was stirred overnight at room temperature then concentrated at 30 °C. Column chromatography (4:1 MeOH-H<sub>2</sub>O) of the residue on Sephadex LH-20 (30 g) gave 22 (25.5 mg, quantitative) as an amorphous mass: [α]<sub>D</sub> -24.0° (*c* 0.6, 1:1 MeOH-H<sub>2</sub>O); <sup>1</sup>H NMR (D<sub>2</sub>O) δ 1.20 (d, 3H, J<sub>5</sub>,6 = 6.6 Hz, H-6b), 2.25 (s, 3H, AcN), 3.33 (t, 1H, J<sub>gem</sub> = J<sub>1</sub>ax,2 = 11.2 Hz, H-1aax), and 4.58 (d, 1H, J<sub>1</sub>,2 = 7.9 Hz, H- 1c). The mass spectrum of 22 (negative ion mode) showed the base peak at m/z 592.5 (M-H)<sup>-</sup>.

O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glyc $ero - \alpha - D - galacto - 2 - nonulopyranosylonate) - (2 \rightarrow 3) - O - (2, 4, 6 - tri - O - benzo$ yl ·  $\beta$  · D · galactopyranosyl) · (1  $\rightarrow$  4) · O · [(2,3,4-tri-O · benzyl ·  $\alpha$  · L · fucopyranosyl)- $(1 \rightarrow 3)$ ]-2-acetamido-1,5-anhydro-6-O-benzoyl-2-deoxy-D-glucitol (23). To a solution of 16 (102 mg, 0.081 mmol) and 17 (80 mg, 0.172 mmol) in benzene (1.5 mL) were added MS-4Å (180 mg) and the mixture was stirred overnight at room temperature then cooled to 0 °C. DMTST (130 mg, 0.486 mmol) was added to the mixture, and this was stirred for 12 h at 15 °C. After completion of the reaction, MeOH (1 mL) was added and the mixture was neutralized with Et3N, and the precipitate was collected and washed with CH2Cl2. A workup as described for 15 gave 23 (120 mg, 88%) as an amorphous mass:  $[\alpha]_D$  +3.3° (*c* 0.8, CHCl<sub>3</sub>); <sup>1</sup>H NMR  $(CDCl_3) \delta 1.00 (d, 3H, J_{5.6} = 6.4 Hz, H-6b), 1.52-2.15 (6s, 18H, 4AcO, 2AcN),$ 2.40 (dd, 1H,  $J_{gem} = 12.5 \text{ Hz}$ ,  $J_{3eq,4} = 4.5 \text{ Hz}$ , H-3deq), 3.78 (s, 3H, MeO), 4.77 (m, 1H, H-4d), 5.13 (d, 1H,  $J_{1,2} = 3.8$  Hz, H-1b), 5.28 (d, 1H,  $J_{6,7} = 2.8$  Hz,  $J_{7,8}$ = 10.0 Hz, H-7d), 5.38 (d, 1H, H-4c), 5.48 (dd, 1H, J<sub>2,3</sub> = 10.1 Hz, H-2c), 5.68 (m, 1H, H-8d), and 7.21-8.20 (m, 35H, 7Ph).

Anal. Calcd for C89H95N2O29 (1673.8): C, 63.86; H, 5.72; N, 1.67. Found: C, 63.71; H, 5.55; N, 1.78.

*O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)-(2 $\rightarrow$ 3)-*O*-(2,4,6-tri-*O*-benzoyl-β-D-galactopyranosyl)-(1 $\rightarrow$ 4)-*O*-[(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-(1 $\rightarrow$ 3)]-2-acetamido-1,5-anhydro-6-*O*-benzoyl-2-deoxy-D-glucitol (24). A solution of 23 (158 mg, 0.095 mmol) in EtOH (20 mL) and AcOH (5 mL) was stirred with 10% Pd-C (160 mg) overnight at room temperature under hydrogen, then filtered and concentrated. The residue was treated with Ac<sub>2</sub>O (1 mL) and pyridine (2 mL) overnight at room temperature and concentrated. Column chromatography (50:1 CH<sub>2</sub>Cl<sub>2</sub>-MeOH) of the product on silica gel (30 g) gave 24 (120 mg, 85%) as an amorphous mass: [ $\alpha$ ]<sub>D</sub> +0.5° (c 1.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.12 (d, 3H, J5,6 = 6.4 Hz, H-6b), 1.49-2.06 (9s, 27H, 7AcO, 2AcN), 2.40 (dd, 1H, J<sub>gem</sub> = 12.5 Hz,  $J_{3eq,4} = 4.5$  Hz, H-3deq), 3.78 (s, 3H, MeO), 4.77 (m, 1H, H-4d), 5.35 (d, 1H,  $J_{1,2} = 3.8$  Hz, H-1b), 5.48 (dd, 1H,  $J_{1,2} = 8.1$  Hz,  $J_{2,3} = 10.1$  Hz, H-2c), 5.65 (m, 1H, H-8d), and 7.26-8.15 (m, 20H, 4Ph).

Anal. Calcd for C74H83N2O32 (1529.5): C, 58.11; H, 5.47; N, 1.83. Found: C, 58.24; H, 5.61; N, 1.59.

*O*-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic Acid)-(2→3)-*O*-β-D-galactopyranosyl-(1→4)-*O*-[α-L-fucopyranosyl-(1→3)]-2-acetamido-1,5-anhydro-2-deoxy-D-glucitol (25). To a solution of 24 (80 mg, 0.054 mmol) in MeOH (2 mL) was added NaOMe (6 mg), and the mixture was stirred overnight at room temperature. Water (1 mL) was added to the mixture, and this was stirred for 5 h at room temperature, neutralized with Amberlite IR-120 (H<sup>+</sup>) resin, and filtered, the resin was washed with MeOH, and the combined filtrate and washings were concentrated. Column chromatography (MeOH) of the residue on Sephadex LH-20 (30 g) gave 25 (42 mg, quantitative) as an amorphous mass: [α]<sub>D</sub> -25.0° (c 0.7, MeOH); <sup>1</sup>H NMR (Me<sub>2</sub>SO-d6) δ 1.12 (d, 3H, J<sub>5,6</sub> = 6.2 Hz, H-6b), 1.89,1.98 (2s, 6H, 2AcN), 3.19 (t, 1H, H-1aax), 4.42 (d, 1H, J<sub>1,2</sub> = 7.6 Hz, H-1c), and 5.02 (d, 1H, J<sub>1,2</sub> = 2.8 Hz, H-1b).

Anal. Calcd for C<sub>31</sub>H<sub>52</sub>N<sub>2</sub>O<sub>22</sub> (804.8): C, 46.26; H, 6.51; N, 3.48. Found: C, 46.15; H, 6.80; N, 3.34.

#### ACKNOWLEDGMENT

This work was supported in part by a Grant-in-Aid (No. 05274102 and No. 06281227) for the Scientific Research on Priority Areas from the Ministry of Education, Science and Culture of Japan.

#### REFERENCES

- 1. M. P. Bevilacqua, S. Stengelin, M. A. Gimbrone, Jr. and B. Seed, Science, 243, 1160 (1989).
- 2. G. I. Johnston, R. G. Cook and R. P. McEver, Cell, 56, 1033 (1989).
- 3. L. A. Lasky, M. S. Singer, T. A. Yednock, D. Dowbenko, C. Fennie, H. Rodriguez, T. Nguyen, S. Stachel and S. D. Rosen, *Cell*, **56**, 1045 (1989).

- 4. T. F. Tedder, C. M. Isaacs, E. J. Ernst, G. D. Demetri, D. A. Adler and C. M. Disteche, J. Exp. Med., 170, 123 (1989).
- J. B. Lowe, L. M. Stoolman, R. P. Nair, R. D. Larsen, T. L. Berhend and R. M. Marks, *Cell*, 63, 475 (1990).
- 6. L. M. Phillips, É. Nudelman, F. C. A. Gaeta, M. Perez, A. K. Singhal, S. Hakomori and J. C. Paulson, *Science*, **250**, 1130 (1990).
- 7. G. Walz, A. Aruffo, W. Kolanus, M. P. Bevilacqua and B. Seed, Science, 250, 1132 (1990).
- 8. E. L. Berg, J. Magnani, R. A. Warnock, M. K. Robinson and E. C. Butcher, Biochem. Biophys. Res. Commun., 184, 1048 (1992).
- 9. D. Tyrrell, P. James, N. Rao, C. Foxall, S. Abbas, F. Dasgupta, M. Nashed, A. Hasegawa, M. Kiso, D. Asa, J. Kidd and B. K. Brandley, *Proc. Natl. Acad. Sci. USA*, **88**, 10372 (1991).
- 10. B. K. Brandley, M. Kiso, S. Abbas, P. Nikrad, O. Srivasatava, C. Foxall, Y. Oda and A. Hasegawa, *Glycobiology*, **3**, 633 (1993).
- M. Larkin, T. J. Ahern, M. S. Stoll, M. Shaffer, D. Sako, J. O'Brien, C.-T. Yuen, A. M. Lawson, R. A. Childs, K. M. Barone, P. R. Langer-Safer, A. Hasegawa, M. Kiso, G. R. Larsen and T. Feizi, *J. Biol. Chem.*, 267, 13661 (1992).
- 12. M. Yoshida, A. Uchimura, M. Kiso and A. Hasegawa, *Glycoconjugate J.*, **10**, 3 (1993).
- 13. A. Hasegawa, T. Ando, M. Kato, H. Ishida and M. Kiso, Carbohydr. Res., 257, 67 (1994).
- 14. T. Terada, M. Kiso and A. Hasegawa, Carbohydr. Res., 259, 201 (1994).
- 15. A. Hasegawa, M. Kato, T. Ando, H. Ishida and M. Kiso, Carbohydr. Res., in press.
- 16. M. Kiso, K. Furui, K. Ando and A. Hasegawa, J. Carbohydr. Chem., 12, 673 (1993).
- 17. A. Hasegawa, A. Uchimura, H. Ishida and M. Kiso, *Biosci. Biotech. Biochem.*, in press.
- P. J. Green, T. Tamatani, T. Watanabe, M. Miyasaka, A. Hasegawa, M. Kiso, C.-T. Yuen, M. S. Stoll and T. Feizi, *Biochem. Biophys. Res. Commun.*, 188, 244 (1992).
- C.-T. Yuen, K. Bezouska, J. O'Brien, M. Stoll, R. Lemonie, A. Lubineau, M. Kiso, A. Hasegawa, N. J. Bockovich, K. C. Nicolaou and T. Feizi, *J. Biol. Chem.*, 269, 1595 (1994).
- 20. R. R. Schmidt and J. Michel, Angew. Chem. Int. Ed. Engl., 19, 731 (1980).
- 21. A. Kameyama, H. Ishida, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, 200, 269 (1990).
- a) P. Fügedi and P. J. Garegg, Carbohydr. Res., 149, c9 (1986); b) M. Ravenscroft, R. M. G. Robert and J. G. Tillett, J. Chem. Soc., Perkin Trans. 2, 1569 (1982).
- 23. A. Kameyama, H. Ishida, M. Kiso and A. Hasegawa, J. Carbohydr. Chem., 10, 549 (1991).
- 24. A. Hasegawa, K. Ito, H. Ishida and M. Kiso, J. Carbohydr. Chem., in press. 25. K. Jansson, T. Freid, J. Kihlberg and G. Magnusson, Tetrahedron Lett., 27.
- 25. K. Jansson, T. Frejd, J. Kihlberg and G. Magnusson, *Tetrahedron Lett.*, 27, 753 (1986).
- a) M. Numata, M. Sugimoto, K. Koike and T. Ogawa, *Carbohydr. Res.*, 163, 209 (1987); b) T. Murase, H. Ishida, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, 188, 71 (1989).
- A. Hasegawa, Y. Hioki, E. Seki, M. Kiso and I. Azuma, Agric. Biol. Chem., 50, 1873 (1986).
- 28. P. J. Garegg, H. Hultberg and S. Wallin, Carbohydr. Res., 108, 97 (1982).